



Specification Sheet

Product:	Drying Buffer/Stabilizer, biotin-free Casein based, ready-to-use
Code:	#DBS-C
Protein concentration :	10 mg/ml
Buffer base:	0,15M PBS pH 7,3
Preservative:	BND, 550 ppm
Lot:	859147
Storage:	+2°C ~ +8°C
Expiration date:	10/2022
Recommended use:	ready-to-use reagent for preparing dried non porous and porous immunosorbents stable real-time at room/ambient temperature

Warm up to room temperature prior to use

In ELISA test kits production DBS-C is applicable in two different ways – with and without wash after coating immunoplate with a capture antibody/antigen.

Protocol 1: exhaustively aspirate coating buffer from immunoplate wells at the end of the antibody/antigen adsorption process and immediately fill them with DBS-C. If immunoplate was coated with e.g. (typically) 100 µl antibody/antigen solution per well, applying DBS-C in the larger volume – 150-200 µl will result in the better NSB blocking. Incubate at least one hour at room/ambient temperature to allow for effective casein adsorption. Longer – up to 24 hours – incubation will aid to the better blocking. Do not wash DBS-C out of the plate, aspirate it as completely as possible immediately before drying. Store dried immunosorbent in water-impenetrable pouch with proven desiccant.

Protocol 2: wash immunoplate with appropriate wash buffer following antibody/antigen adsorption. Aspirate rests of the wash buffer as completely as possible and then apply DBS-C. Further steps are identical to Protocol 1.

On High Binding Capacity plates (capable of binding 400-500 ng IgG/cm²) coated with IgG/F(ab')₂ and/or peptide antigens in concentration about 1 µg/ml both protocols may work equally well, i.e. Protocol 2 with wash will have no advantage over Protocol 1 without wash. Rationale behind is that 1 µg/ml at 100 µl per well translates into 100 ng per well – this amount of the applied material will most likely be almost completely adsorbed during 18-24-hour coating procedure.